

# **Nuclear microprobe facility at iThemba LABS in multidisciplinary applications – key elements of a success story**

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## **Background:**

(from IAEA technical meeting, Cape Town, 7-10 November 2005)

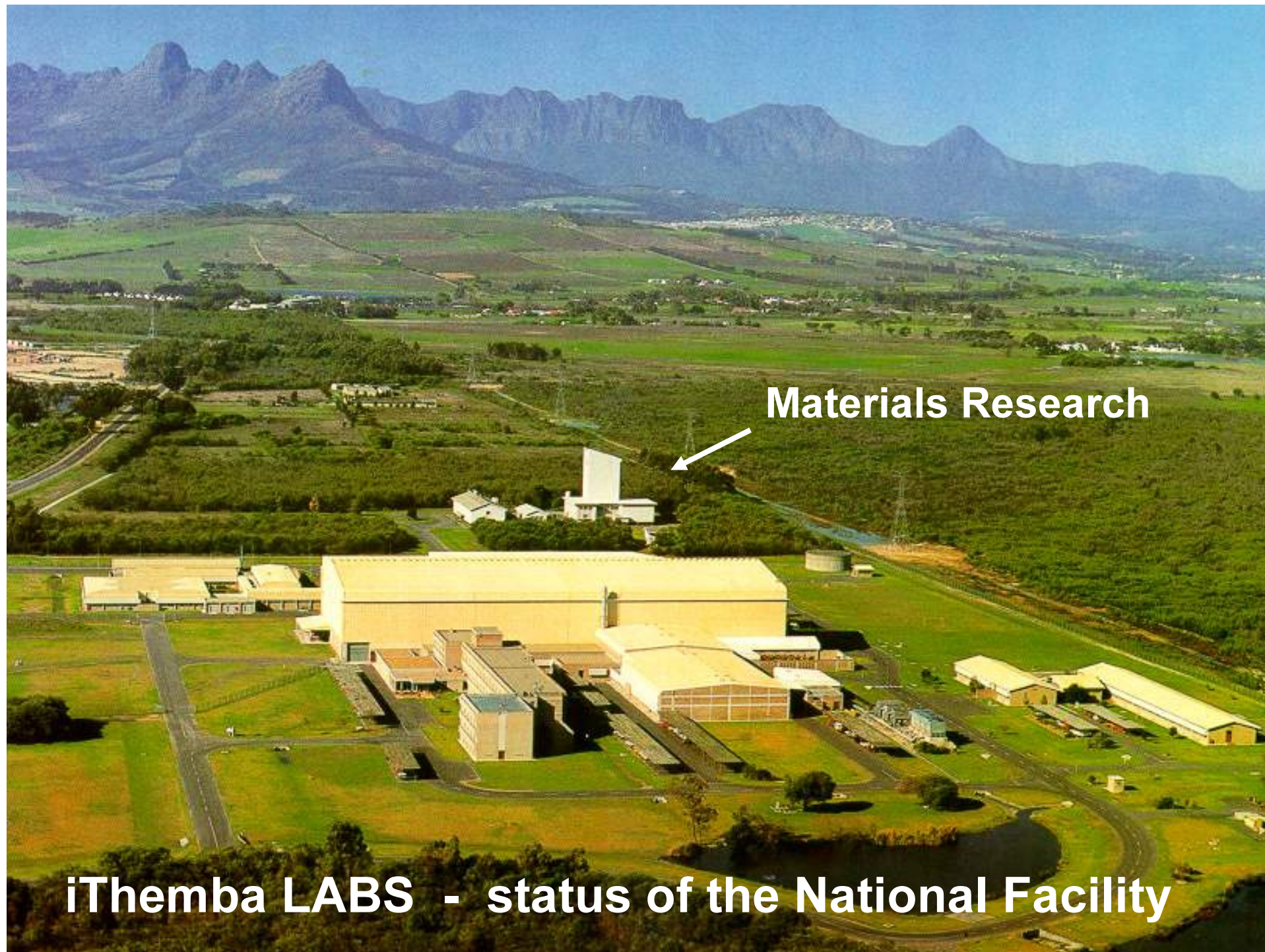
**“Accelerator-based physics has grown from its roots in nuclear physics research into a mature broad ranging science and technology serving social and economic development**

**Innovations in accelerator technologies, techniques, and nuclear instrumentation are continuing to enable successful solutions to many challenging problems,**

**both in the traditional application areas of engineering and physical sciences and increasingly, in the areas of environmental and life sciences.”**

**This is**

**well reflected in usage of our van de Graaff accelerator at Materials Research Department of iThemba LABS**



**Materials Research**

**iThemba LABS - status of the National Facility**



# **Basic requirements to reach disciplines other than physics and to make accelerator-based techniques competitive:**

## **Reliable accelerator operation**

- Operators** (maintenance + standby availability)
- Long-term stability, small terminal voltage ripple

## **Proper equipment**

- Permanent setup (electronic modules, measurement geometry)
- Data acquisition suitable for the needs (flexible but easy to use)
- Best available software for data processing

## **Dedicated, experienced scientist(s) (analysts)**

## **Auxiliary equipment available on-site**

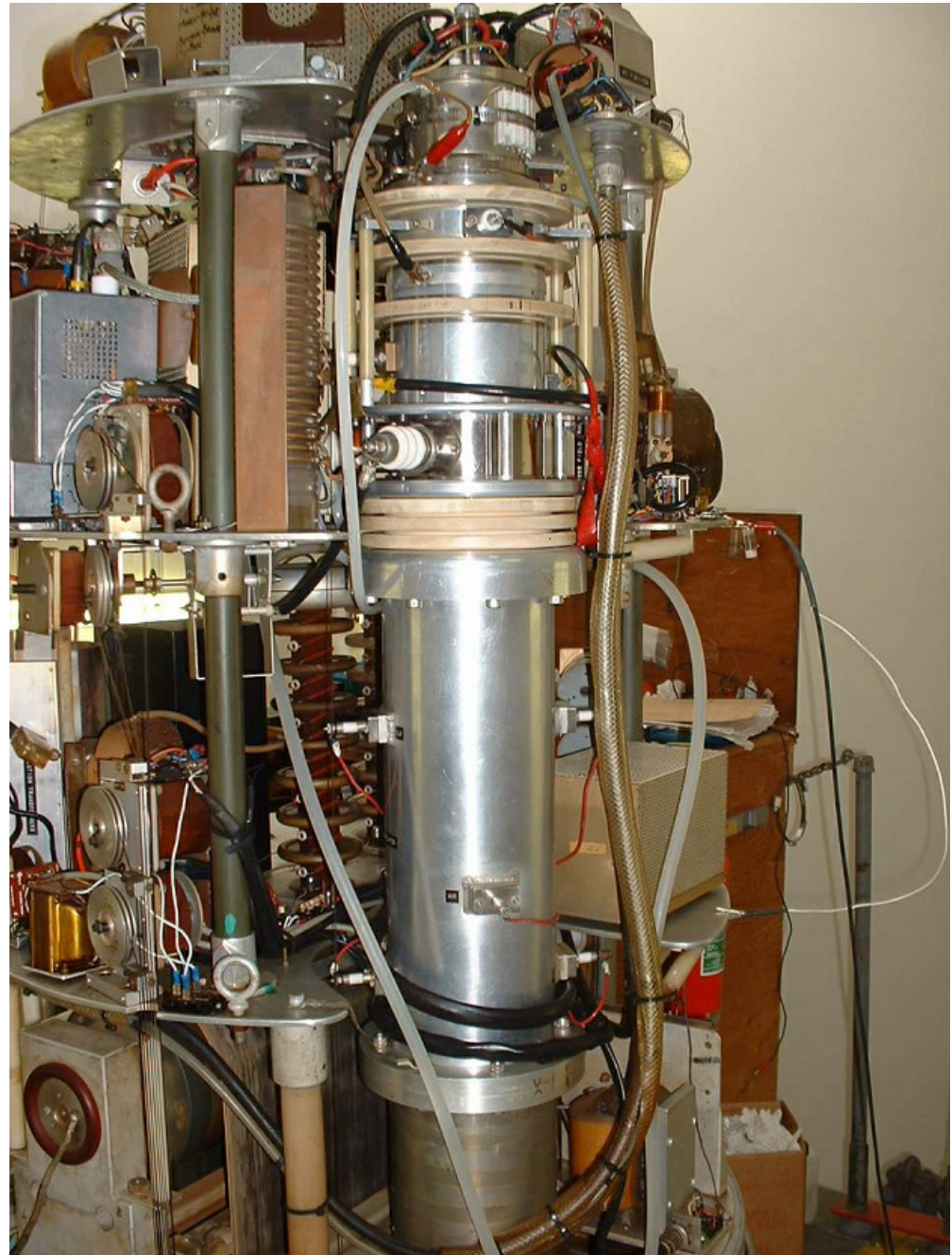
**Cryo-preparation laboratory and expertise in specimen preparation**



# 6 MV CN Van de Graaff Accelerator

- Manufactured by High Voltage Engineering – USA.
- Installed in 1963
- Vertical machine, Voltage range: 0.6 to 6 MV
- Tank 18 ton. Pressure: 16 Bar, Gas: N<sub>2</sub>=80% and CO<sub>2</sub>=20%
- Gas compressor, dryer and storage tank.
- Opening cycle 27 hours
- Current: DC 20 uA max. 1.5 nS, Pulsed @ 2 MHz 5 uA
- Drive motor 10 kW 60 Hz 1700 RPM, Belt speed 60 km/h
- Terminal generator 115 V 400 Hz 2 kW max
- Column/accelerating tube: 2 tubes with 132 insulated rings in total, with a resistor chain of  $10^{11}$  ohm

The high-voltage  
terminal of the  
accelerator with the  
dome removed



## **Development of the Van de Graaff accelerator and beam lines**

**Courtesy: L. Conradie, Accelerator Department, iThemba LABS**

Replacement of the 30 year old resistor chain

Replace the top section of the acceleration tube

Installation of 8 new steerer magnets

Installation of 8 new Faraday cups along the beam lines

Installation of improved energy stabilization slit system

Increase the apertures of the existing quadrupole magnets

Installation of a complete vacuum control system

Installation of a computer controlled safety interlocking system for the protection of staff and equipment

Computer control system for all the power supplies and slits of the beam lines





Van de Graaff  
terminal-voltage  
stabilizing unit



The newest Terminal Potential Stabilizer - NEC TPS 6.0

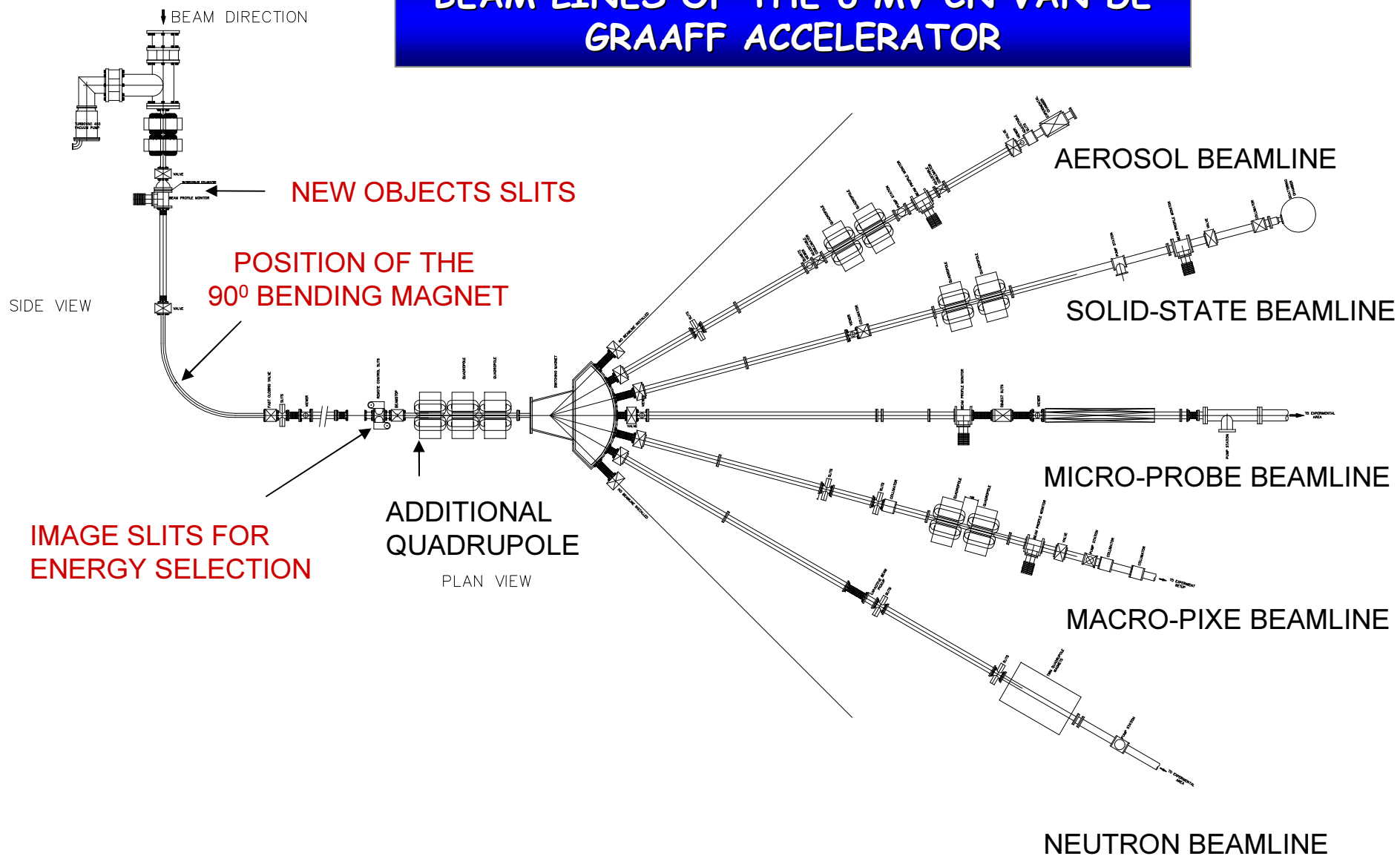


Beam ripple now  
~ 900 V at 3 MV

Was ~ 1400 V

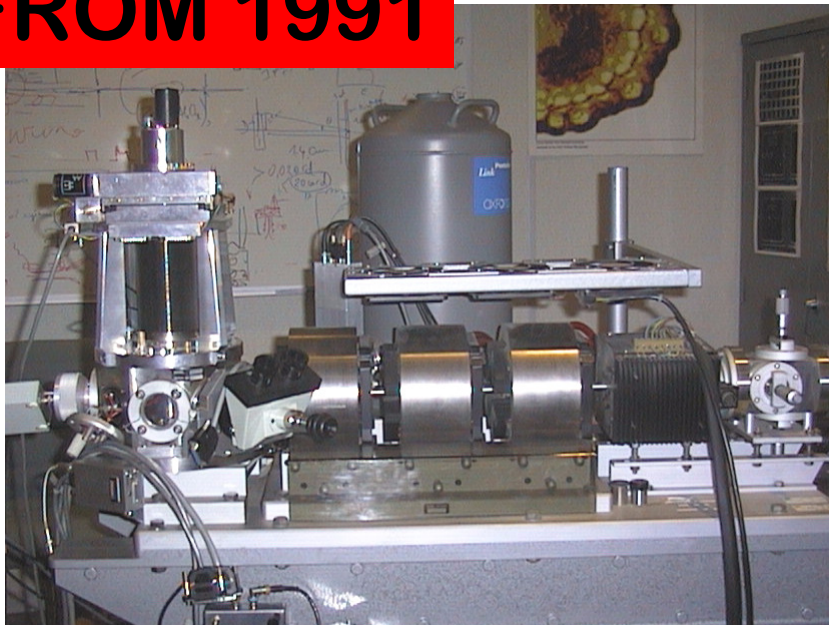
VAN DE GRAAFF ACCELERATOR

# BEAM LINES OF THE 6 MV CN VAN DE GRAAFF ACCELERATOR





# FROM 1991

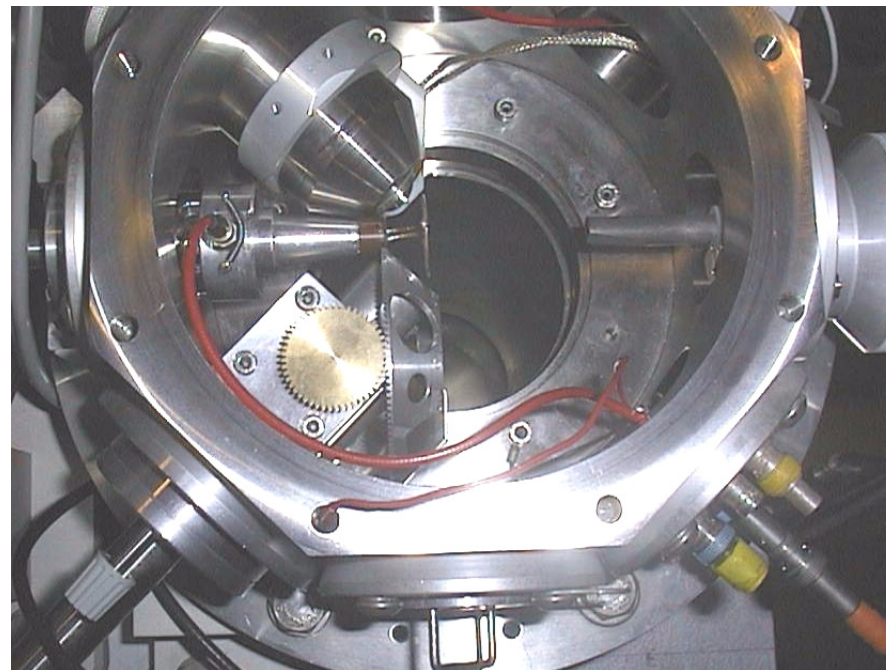


## Features:

- OM150 Triplet and scanning system (Oxford Microbeams) Best beam spot at 100 pA  
X – 1.6  $\mu\text{m}$  ; Y – 0.6  $\mu\text{m}$   
**Typical sizes used here X – 3  $\mu\text{m}$  ; Y – 3  $\mu\text{m}$**
- Chamber modified on-site with stepper motors for automated specimen movement;
- permanently mounted set of standards:  
44 pure metals, 53 minerals, fused glasses
- On-demand beam deflection system  
(loop time ca. 900 ns)

## Inside the NMP chamber:

- Si(Li) detector 80 mm<sup>2</sup> and wheel with filters or HPGe detector 100 mm<sup>2</sup>
- Annular Si surface –barrier detector for backscattering analysis (BS, RBS)
- Microscope (specimen viewed at 45°)
- Faraday cup and PIN diode for on/off axis STIM
- Channeltron for Secondary Electron imaging





## **Beam size:**

300 nm for 100 pA proton beam current  
(best laboratories)

100 pA is considered as low current limit for analytical work  
– PIXE, (R)BS, NRA, PIGE

100-200 nm for low current techniques (STIM, techniques  
based on single ion hits)

## **Our present performance:**

**2.6  $\mu\text{m}$  x 3  $\mu\text{m}$  for 100 pA protons (3 MeV)**

**1.2  $\mu\text{m}$  x 2.3  $\mu\text{m}$  for 100 pA alpha particles (2 MeV) short**

**2.6  $\mu\text{m}$  x 2.2  $\mu\text{m}$  for 100 pA long run (overnight)**

## Techniques utilizing focused beam:

Combination **PIXE-RBS** used most often;  
results in complementary information

**PIXE** – very easy identification of elements Na – U down to ppm level,  
but limited depth information

**(R)BS** - very good depth information;  
identification of the layer thickness,  
limited to major and minor elements;  
identification of light elements (C, O, N);  
best material: high Z layer on low Z substrate

**ERDA** - Hydrogen analyses in materials at lateral resolution down  
to 30-50 micrometers

**NRA** - selective analysis of light elements and their isotopes  
(e.g. B, Li, F), depth profiling;

**PIGE** - sensitive for F (down to tens of ppm); also B, Li

### **Data acquisition:**

XSYS software on VAX station 4000/VLC acquiring data over ETHERNET LAN from VME/CAMAC front end

### **Processing of PIXE spectra and elemental mapping:**

DEC 2300 Alpha Server running VMS and GEOPIXE-I software

**True elemental images** obtained ON-LINE using DYNAMIC ANALYSIS

#### **OLD**

List-mode files possible  
but **SELDOM USED**  
GEOPIXE-I software  
on Alpha Server

**Quantitative maps  
from scanned areas**

#### **PRESENT**

List-mode files **ALWAYS USED**  
GEOPIXE-II software  
on PC and laptop if needed

**Quantitative maps with all new features**  
+ PIXE, BS, PIGE  
extracted from selected areas



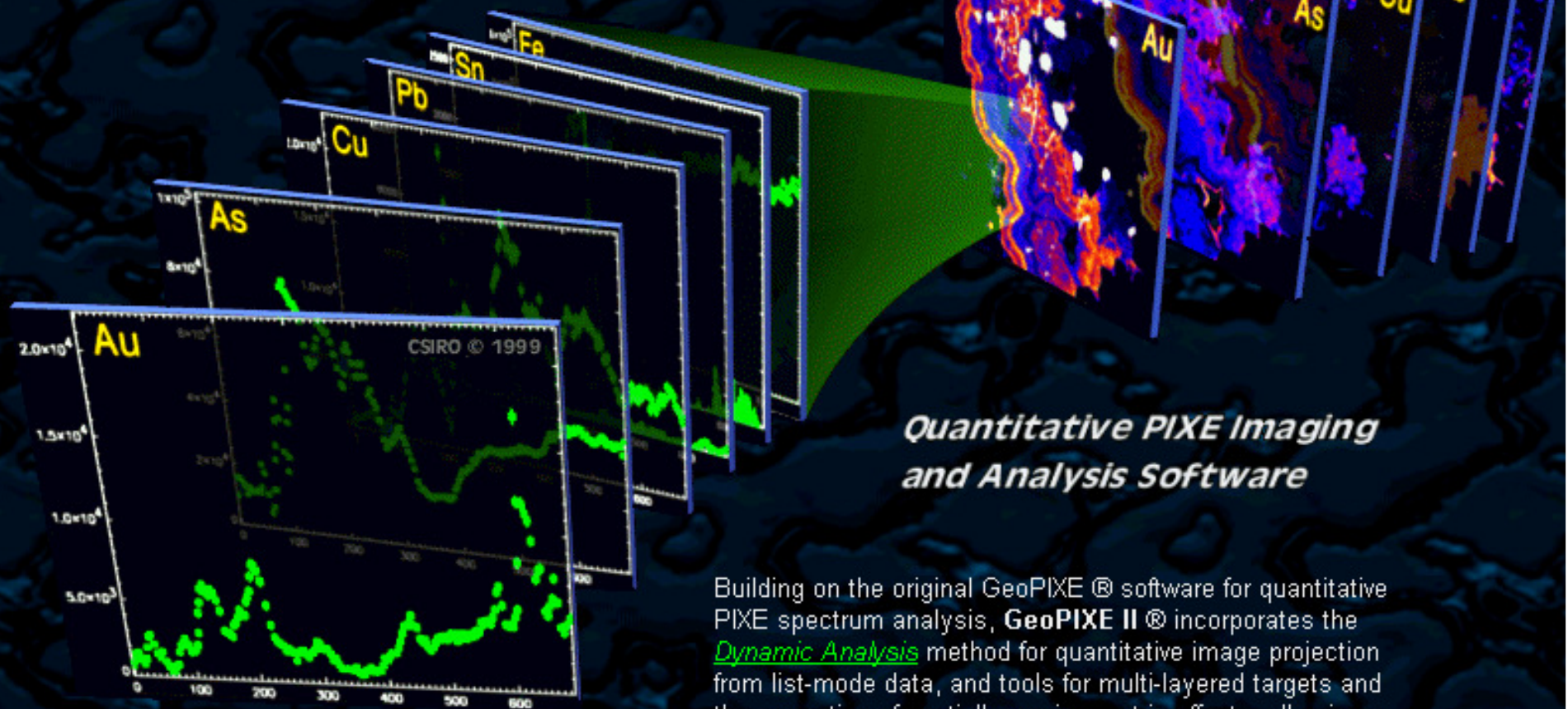
## Processing of list-mode data

<http://www.syd.dem.csiro.au/research/hydrothermal/chris/>

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# GeoPIXE II



*Quantitative PIXE Imaging  
and Analysis Software*

Building on the original GeoPIXE © software for quantitative PIXE spectrum analysis, **GeoPIXE II** © incorporates the [Dynamic Analysis](#) method for quantitative image projection from list-mode data, and tools for multi-layered targets and the correction of spatially varying matrix effects, all using a new efficient PC graphical user interface. [Enquiries welcome.](#)

## Some features:

Data sorting from all detectors (not only PIXE but PIXE+BS+PIGE+...)

Spectrum fit generates **Dynamic Analysis** transform matrix

Projection of EVT data onto quantitative elemental images

- resolves element overlaps
- rejects artefacts from overlapping elements, detector response effects (escape peaks, tails)
- subtracts background
- **groups of lines (K, L) used for construction of images**
- treatment of pileups:
  - using spectrum
  - using images (the only software with this option?)**

Extraction of concentration data from selected “regions”

# (R)BS fitting: RUMP with cross-sections from SigmaCalc

## We rely on this development!

[//www-nds.iaea.org/sigmacalc/](http://www-nds.iaea.org/sigmacalc/)



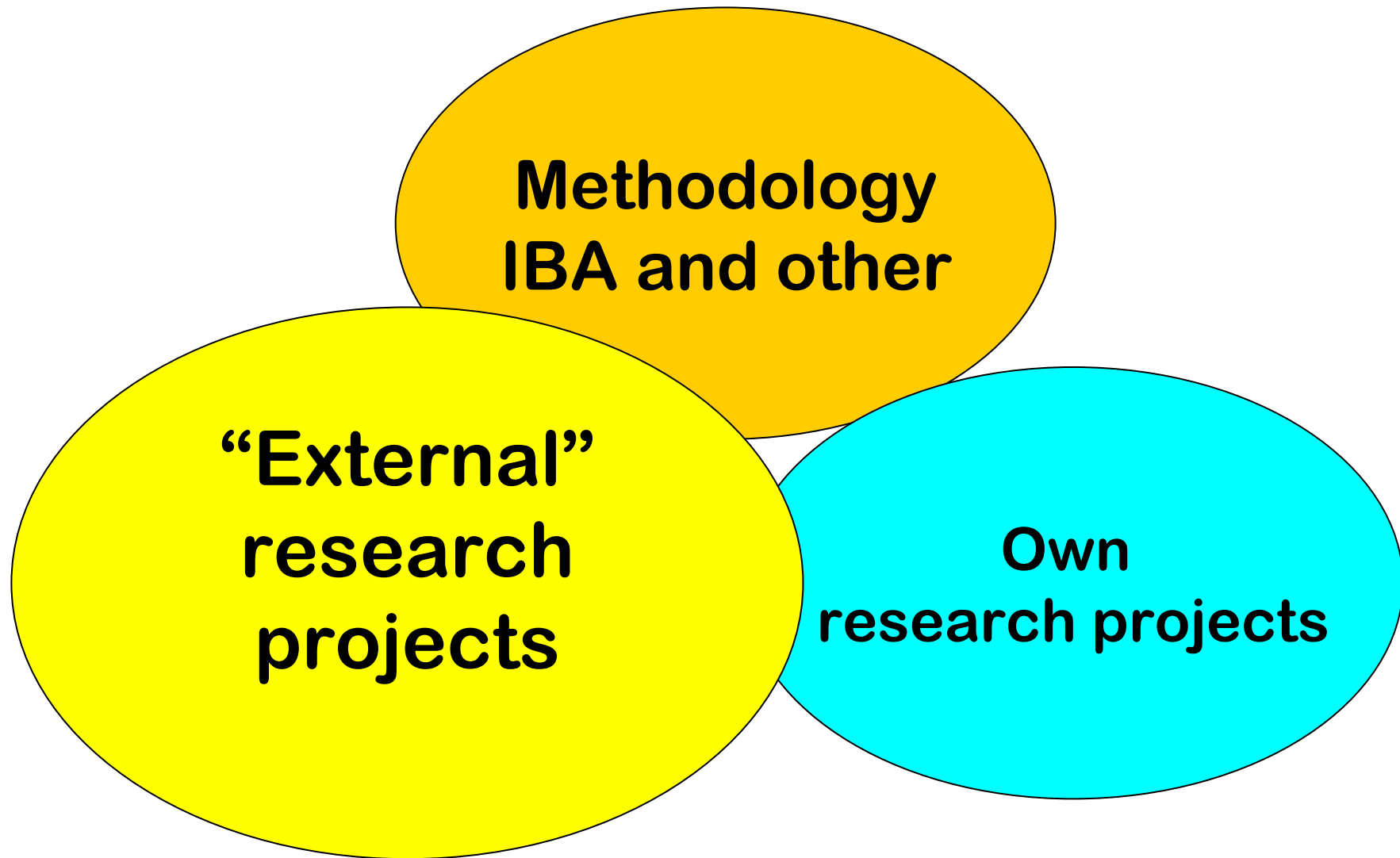
# SigmaCalc

This service provides evaluated (recommended) differential cross sections for Ion Beam Analysis

Format	Element	Reaction	Scattering Angle	About
<input type="checkbox"/> R33	Al-27	<input checked="" type="radio"/> (p,p)	176	<b>General</b>
<b>Energy</b>	H-1	<input type="radio"/> ( $^4\text{He}, ^4\text{He}$ )		Introduction
<input checked="" type="radio"/> keV	He-4	<input type="radio"/> ( $^4\text{He}, p$ )		Calculations
<input type="radio"/> MeV	C-12			Disclaimer
<b>Cross section</b>	N-14			<b>Evaluation</b>
<input checked="" type="radio"/> RTR	O-16			Evaluation procedure
<input type="radio"/> mb/sr(lab)	Al-27			<b>Cross sections</b>
<input type="radio"/> mb/sr(c.m.)	Mg-nat			$^1\text{H}(^4\text{He}, p)$
<input type="radio"/> b/sr(lab)	Si-nat			$^4\text{He}(p, p)$
<input type="radio"/> b/sr(c.m.)	S-nat			$^{12}\text{C}(p, p)$
<b>Delimiter</b>	Country			$^{12}\text{C}(^4\text{He}, ^4\text{He})$
<input checked="" type="radio"/> Tab	E-mail			$^{14}\text{N}(p, p)$
<input type="radio"/> Space				$^{16}\text{O}(p, p)$
<input type="radio"/> Comma				$^{\text{nat}}\text{Mg}(p, p)$
<input type="radio"/> Colon				$^{27}\text{Al}(p, p)$
<input type="radio"/> Semicolon				$^{\text{nat}}\text{Si}(p, p)$
				$^{\text{nat}}\text{S}(p, p)$

*Skip filling the form if you do not wish to receive the information on updates of SigmaCalc*





# Accurate and precise elemental microanalysis is required

Analytical techniques  
with NO spatial resolution

AES  
AAS  
AFS  
ICP-AES  
ICP-MS  
XRF  
TXRF  
PIXE

A microanalytical  
technique  
**Micro-PIXE**

Typically  
Mass needed 1 mg or more

Electron microscopy  
Morphological studies with  
excellent  
spatial resolution

but

Sensitivities of EDS, WDS  
**not sufficient** for studies  
of elemental balances  
of minor and trace elements



# Main Microanalytical Techniques for Imaging – Quantification – Speciation of Chemical Elements in Cells

ELEMENTAL MICROANALYSIS	Analytical depth	Limit of detection	Spatial Resolution	Selectivity	Results	Biological samples
Electron Microscope	0.1 to 1 $\mu\text{m}$	EDS 100 to 1000 $\mu\text{g/g}$	0.1 to 1 $\mu\text{m}$	Multielement	Semi-Quantitative	Dehydrated Hydrated, frozen
	0.05 to 0.1 $\mu\text{m}$	EELS, 100 to 1000 $\mu\text{g/g}$	0.001 $\mu\text{m}$	Multielement, chemical species	Semi-Quantitative	Dehydrated (thin sections)
<b>Nuclear Microprobe</b>	<b>10 to 100 <math>\mu\text{m}</math></b>	<b>1 to 10 <math>\mu\text{g/g}</math></b>	<b>1 <math>\mu\text{m}</math></b>	<b>Multielement</b>	<b>Quantitative (PIXE + RBS)</b>	Dehydrated <b>Hydrated, frozen</b>
Synchrotron Radiation Microprobe	> 100 $\mu\text{m}$	0.1 $\mu\text{g/g}$	0.1 to 1 $\mu\text{m}$	Multielement, chemical species	Semi-Quantitative	Dehydrated Hydrated, frozen
Secondary Ion Mass Spectrometry (SIMS)	0.1 $\mu\text{m}$	< 0.1 $\mu\text{g/g}$	0.05 $\mu\text{m}$	Multielement (isotopes)	Quantitative (Dynamic mode)	Dehydrated (thin sections)
Fluorescence Microscopy	Cell depth	0.01 $\mu\text{g/g}$	1 $\mu\text{m}$	Fluorophores (Na, Mg, Cl, K, Ca, Zn)	Quantitative	Living cells

Courtesy: R. Ortega (slide modified)

## **Facility available free of charge:**

**This big advantage can be lost if no true interest in collaboration exists on both sides**

## **How to make interdisciplinary research productive:**

- Try to identify needs**

  - Research seminars at users' work place**

  - Pilot experiments**

  - Earlier results and publications**

- Project proposals and initial discussions**

- Collaborators from other disciplines present and active during measurements; co-supervision of students**

- Very strong emphasis on data evaluation and reporting  
(do not leave your collaborators with the results, be involved in the discussions, conference presentations, publications)**

# **Applications**

## **(1) Characterisation of materials**

**thin films, surface coatings etc.**

**materials crucial for SA research and industry**

**e.g. related to catalytic converters,**

**water purification,**

**Solar cells, superconducting microelectronics**

**Characterisation of nanomaterials at “micro-scale”**

## **(2) Geology and Ore Processing**

**Elemental mapping capabilities remain an advantage**



# Micro-PIXE and LA-ICP-MS

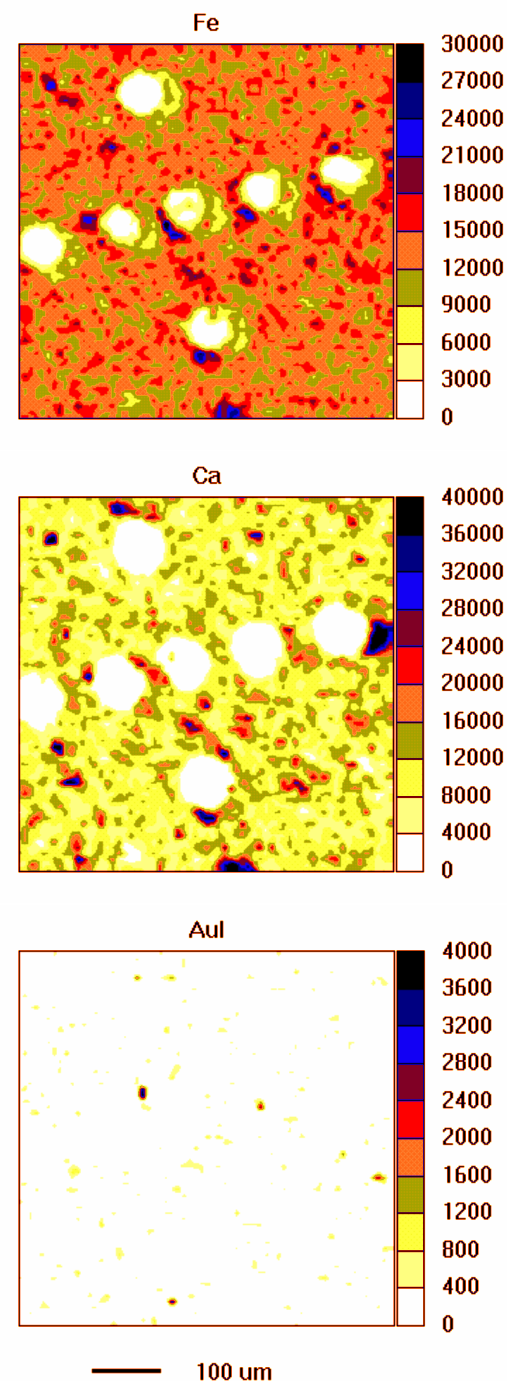
G. Stevens, W. Przybylowicz, L. Martin

X-ray Spectrometry 2004, 33: 216-221.

Studies of gold transport in granitoid  
and silica-undersaturated magmas

Initial analyses of the samples using **LA-ICP-MS** indicated that gold was dissolved within all of the glasses analysed. However, due to the destructive cratering of the technique (figure), the gold content is either diluted (by silicate and oxide presence) or concentrated (where gold crystals are incorporated).

The true required Au content is that  
within the melt alone.



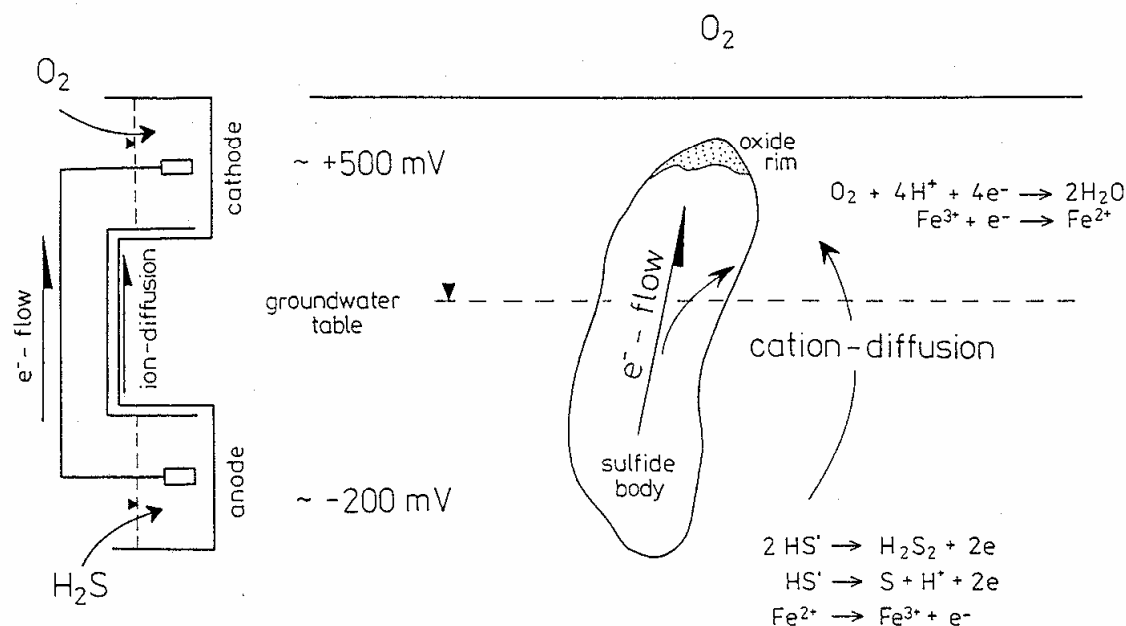
## **One example when true elemental mapping is crucial**

Electrochemical accumulation of visible gold on pyrite and arsenopyrite surfaces

P. Möller and G. Kersten, Mineral. Deposita 29, 404-413 (1994)

In galvanic cell arrangements **gold** is electrochemically deposited on semiconducting sulphide minerals (pyrite, arsenopyrite, chalcopyrite) from aerated as well as H<sub>2</sub>S-saturated, gold-bearing 1M KCl solutions.

Observed cell potential differences (0.4 - 0.6 V) are comparable with known “self-potentials” of natural sulphide ore bodies.



**Fig. 9.** Comparison of electrochemical reactions at a sulfide ore body and an experimental setup of a galvanic cell. Indicated chemical reactions might only be the most important ones in controlling the potential differences of about 700 mV

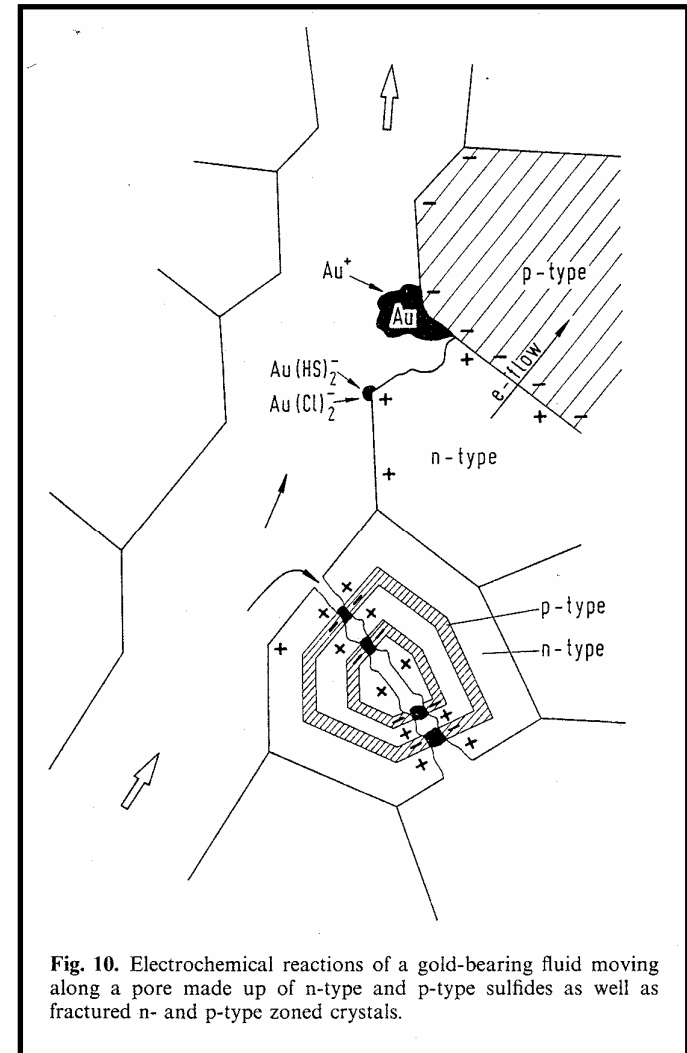
**Gold** preferentially accumulates on the cathode, i.e. under oxidizing conditions.

Galvanic cells simulate conditions occurring at surfaces of chemically inhomogeneous single crystals (zonated).

Sulphide minerals show either **n- or p- type conductivity**. **Visible gold** is accumulated on individual domains of sulphide surfaces that act as **cathodes, i.e. p-type conductors** in n-p junctions.

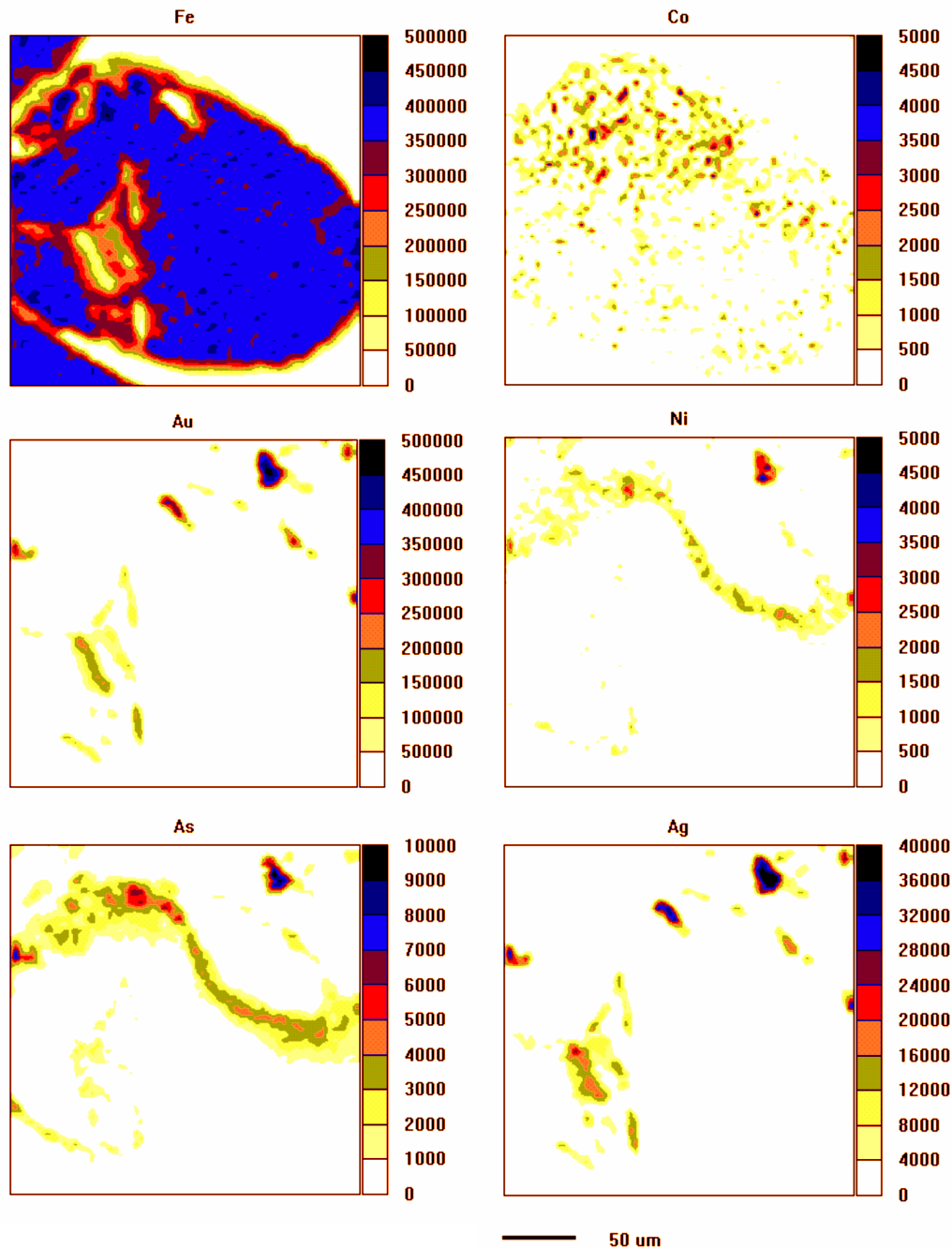
**Arsenic** is the most important element in establishing **p-type conductivity** of pyrite and arsenopyrite.

This explains why **As** is such a powerful pathfinder in gold exploration.



**Fig. 10.** Electrochemical reactions of a gold-bearing fluid moving along a pore made up of n-type and p-type sulfides as well as fractured n- and p-type zoned crystals.

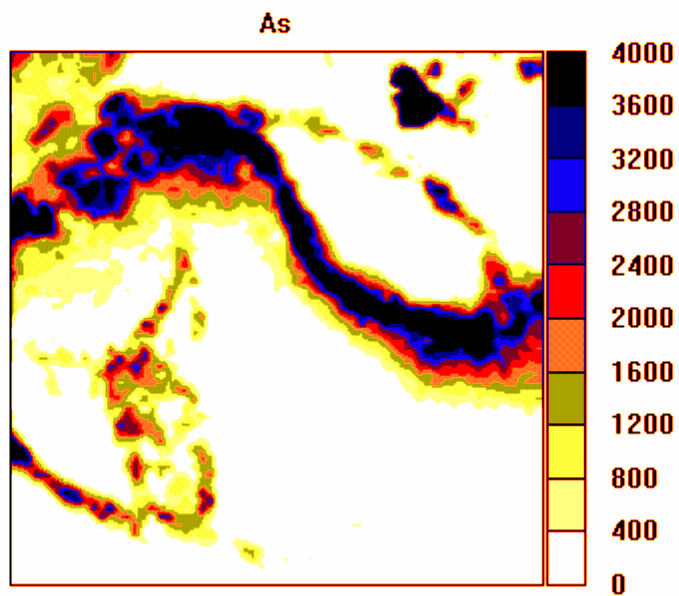
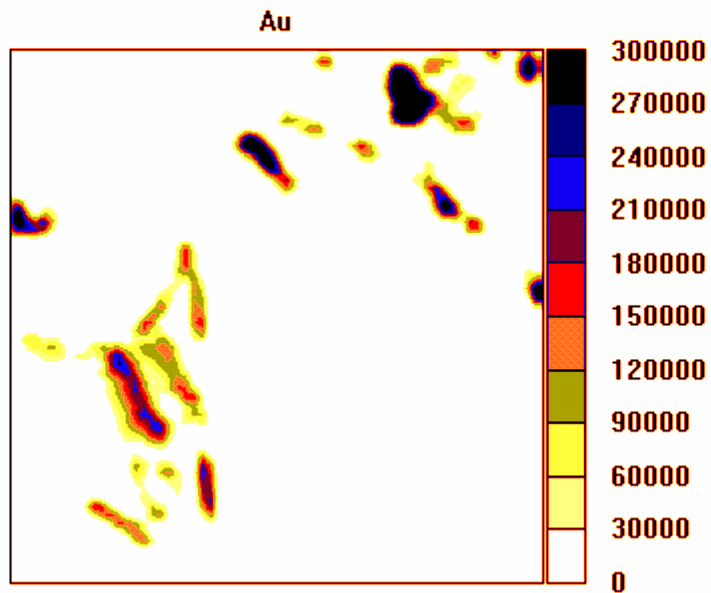




## True elemental maps.

Pyrite grain from the Steyn  
facies of the Basal Reef,  
Unisel Gold Mine in the  
Welkom Goldfield.

3 MeV protons,  
absorber 160 µm Al and  
75 µm Kevlar window

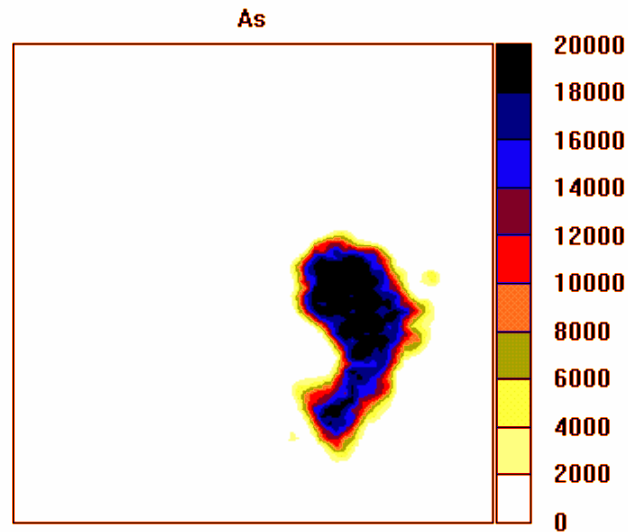


50  $\mu\text{m}$

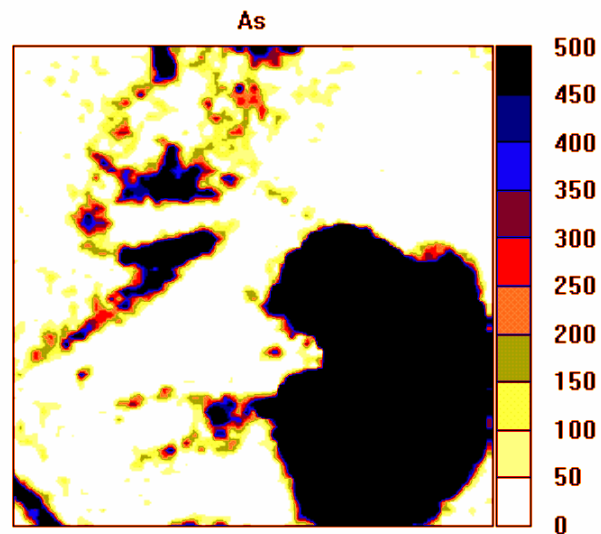
**True elemental maps.**

Au - As relation.

Reduced scale of concentration.



True elemental maps of As zonation in two round, compact pyrite grains from Witwatersrand reef.



Additional details can be distinguished after reduction of the concentration scale from 20000 ppm to 500 ppm.

— 50 um

W.J. Przybylowicz et al. Nucl. Instr. and Meth. B104 (1995) 450-455.

### **(3) Biology and environment; medicine**

unique combination of techniques  
related to specimen preparation and microanalysis  
at ONE research centre

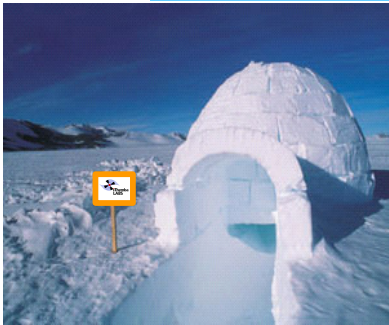
Active long-term research program and established collaborations



# **Where a nuclear microprobe can be applied?**

- **Elemental transport and accumulation**
- **Micro-nutrient uptake and their function in metabolism**
- **Studies on the function of trace elements**
- **Elemental deficiency and toxicity**
- **Environmental pollution**

# Equipment available in the iThemba cryo-laboratory



## Cryo-fixation

### • HPF – High Pressure Freezer

- Leica EM CPC-Cryoworkstation

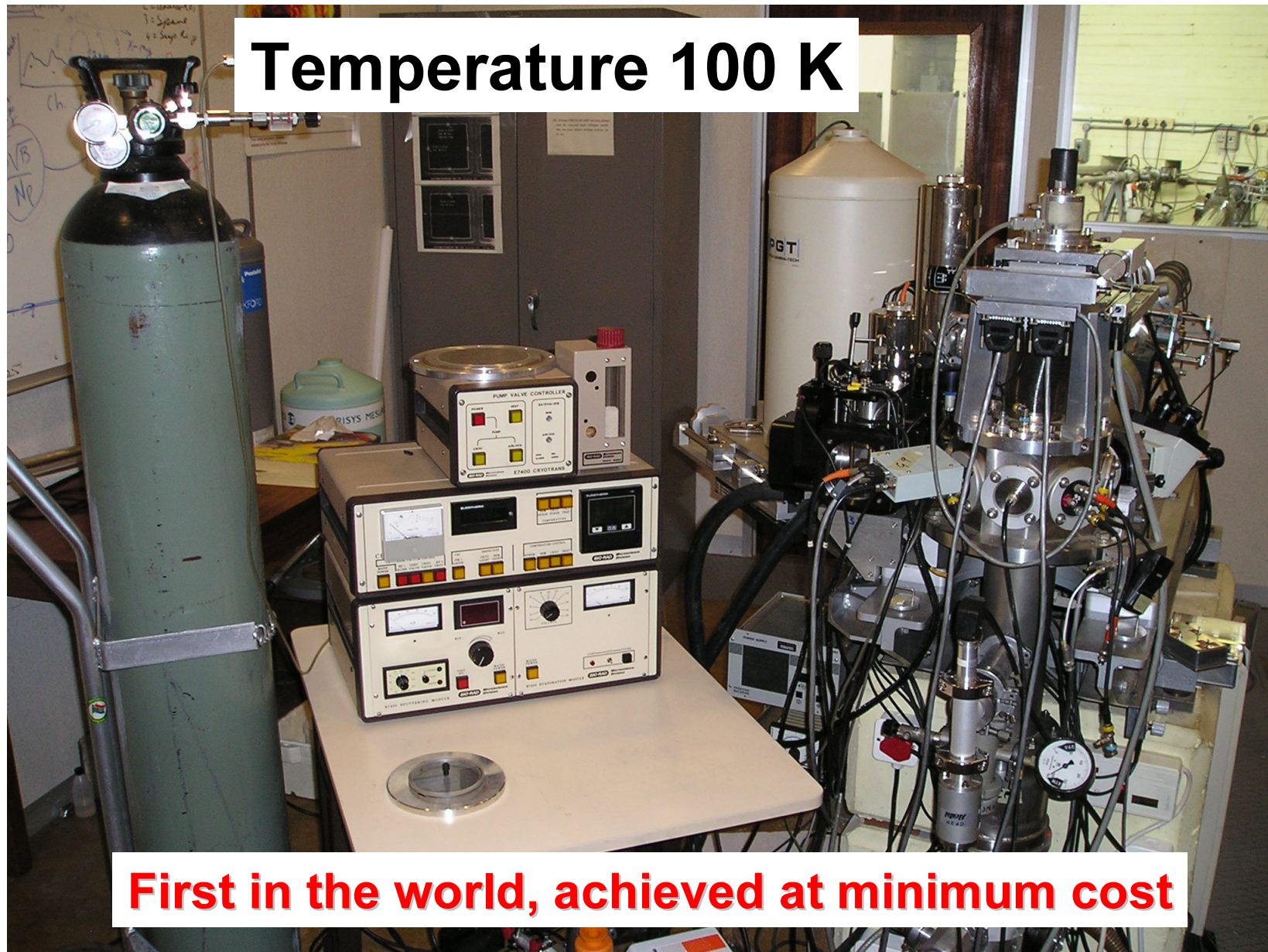


## Further cryo-preparation

- Leica EM CFD Cryosorption Freeze Dryer
- Leica EM AFS Automatic Freeze Substitution
  - Cryo-ultramicrotome
- Dewars for long term storage



Present – with cryotransfer (frozen hydrated specimens)





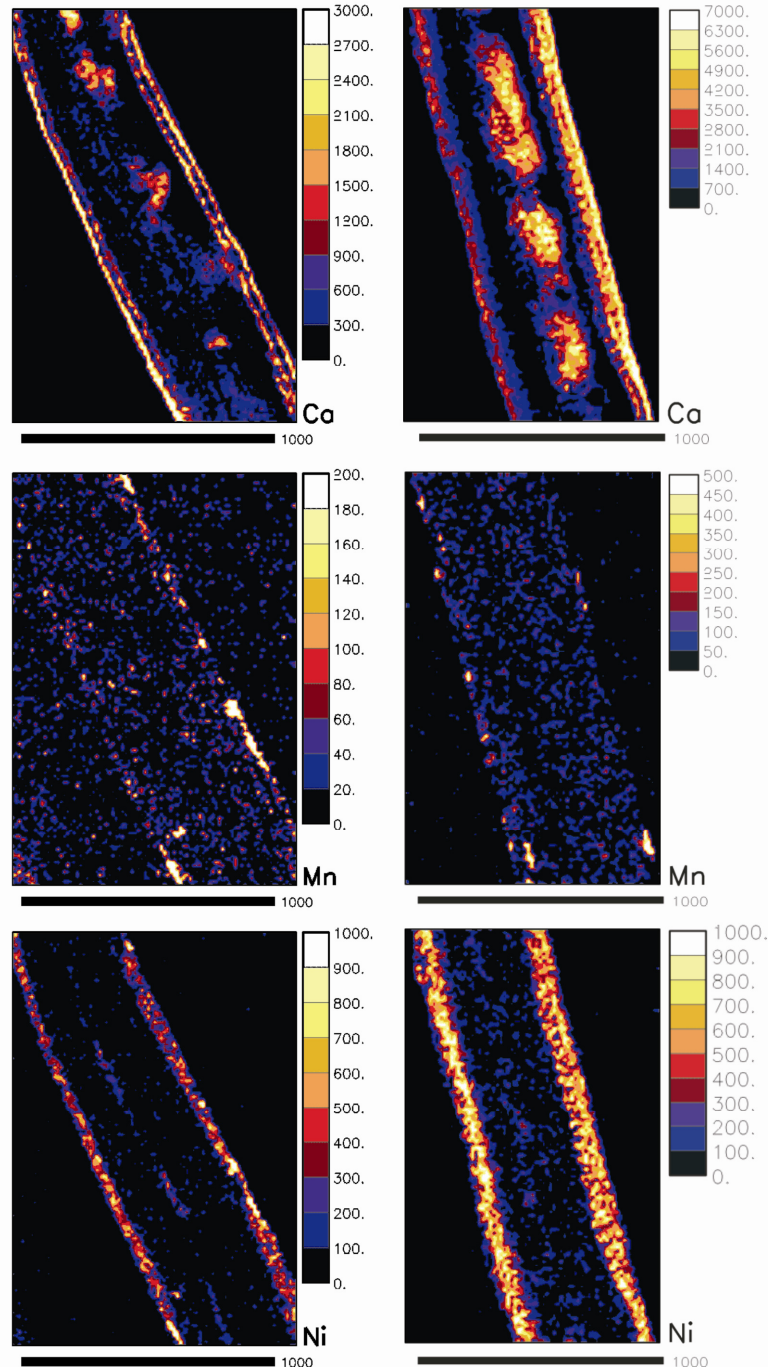
# Comparison of distribution in frozen-hydrated and freeze-dried leaf cross-section

*Senecio anomalochrous*

FH

FD

Tylko G. , Mesjasz-Przybyłowicz J.,  
Przybyłowicz W.J.  
Microsc. Res. Techn. (2007) 70: 55-68.

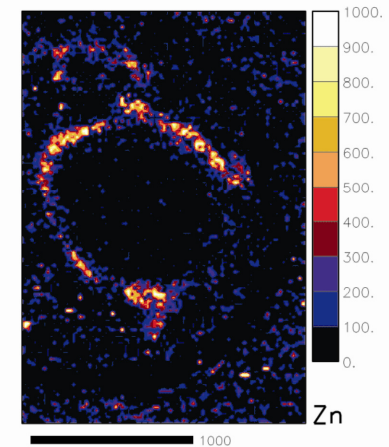
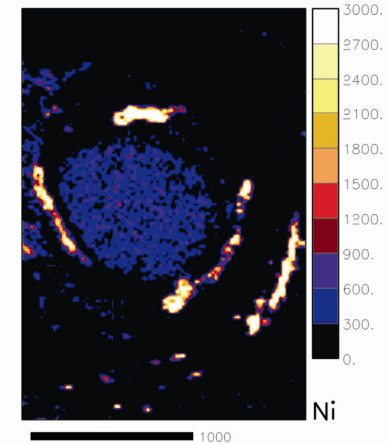
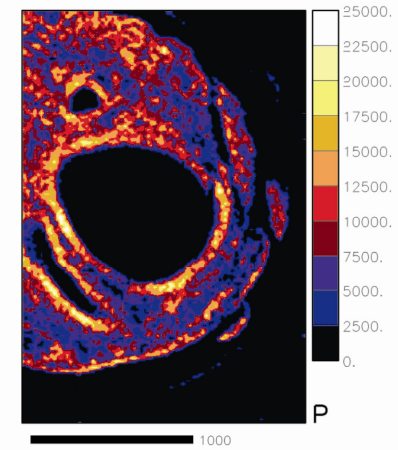
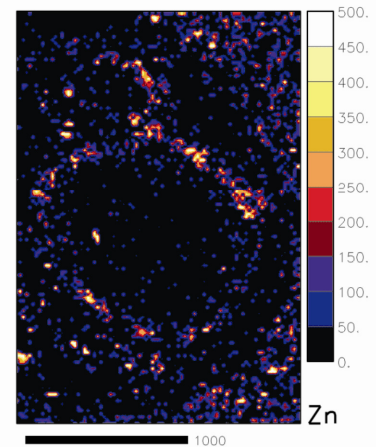
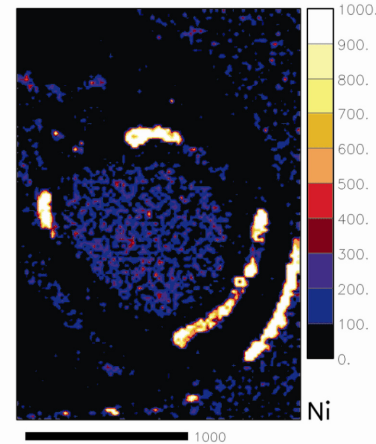
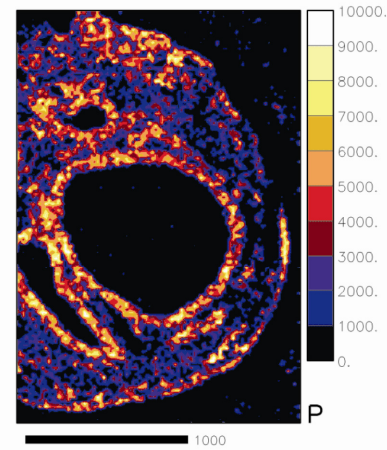




# *Chrysolina pardalina* larvae

## Comparison of distribution in frozen-hydrated and freeze-dried abdominal tissue

FH

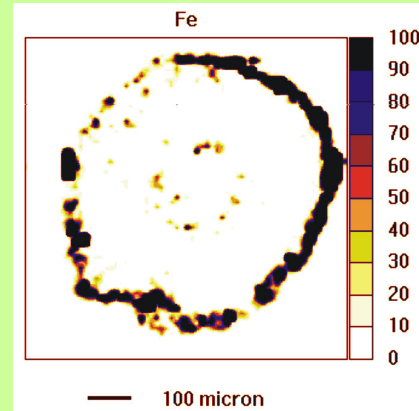
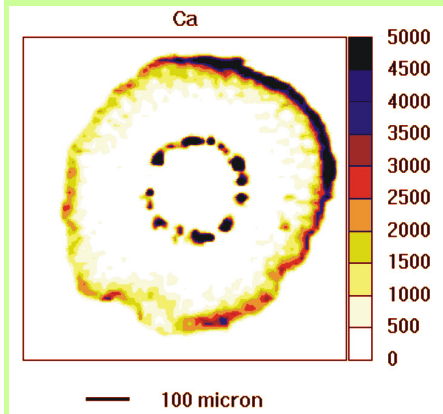


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Tylko G. , Mesjasz-Przybyłowicz J.,  
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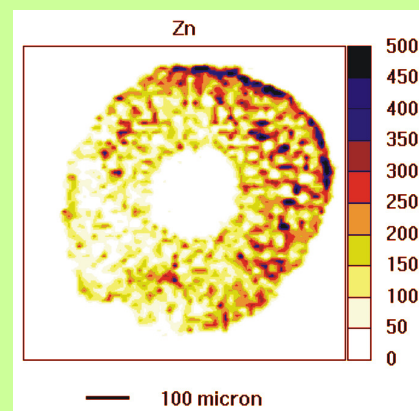
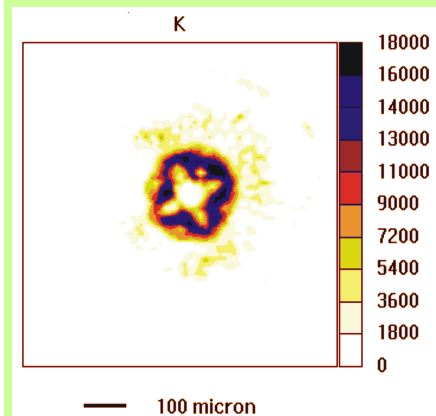


# Studies of the mechanism of Zn uptake in *Phaseolus vulgaris*



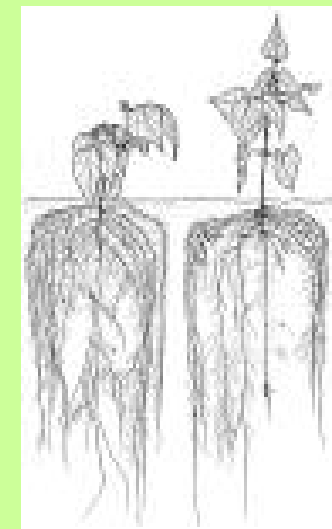
- Zinc is one of the essential micronutrients and an important component of a number of enzymes.

- The results clearly indicate **higher concentrations of zinc in the cortex**



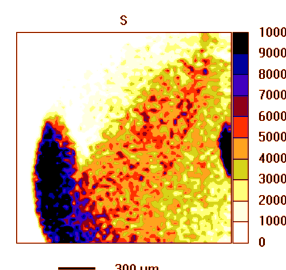
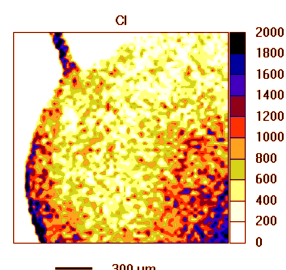
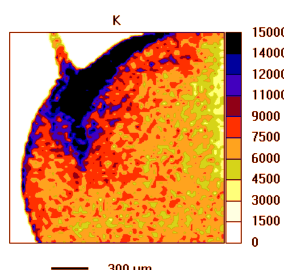
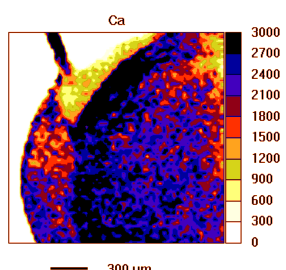
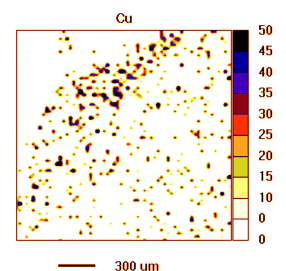
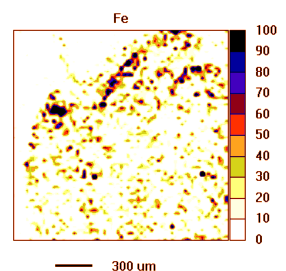
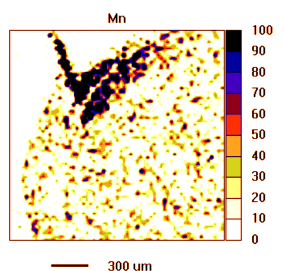
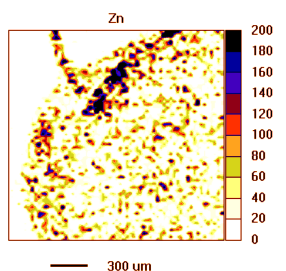
- **Zinc** is not transported to the xylem due to a **passive absorption mechanism**.

- In contrast, **potassium** was mainly located inside the endodermis which may be an evidence for an **active transport across the endodermis** into the conducting tissue of the xylem



J. A. van As, J.H. Jooste, J. Mesjasz-Przybylowicz, W.J. Przybylowicz.  
Proc. 34<sup>th</sup> Conf. Electron Microsc. Soc. South Afr. 25 (1995) 34.

# Studies of elemental distribution in seeds of *Biscutella laevigata* from zinc wastes in Olkusz, Poland

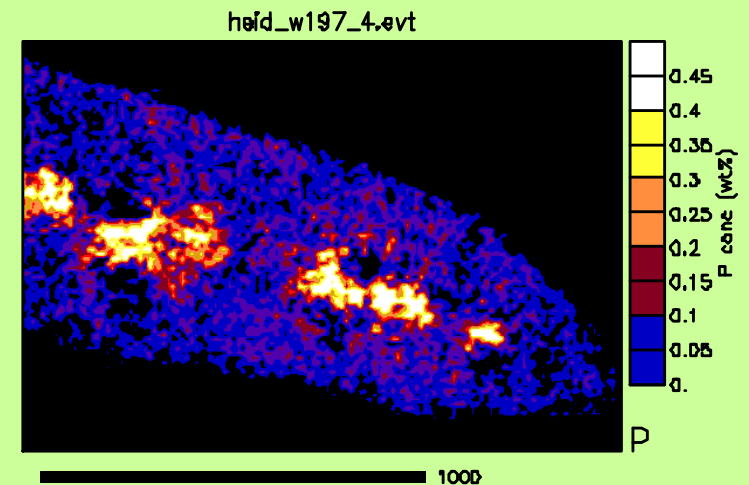
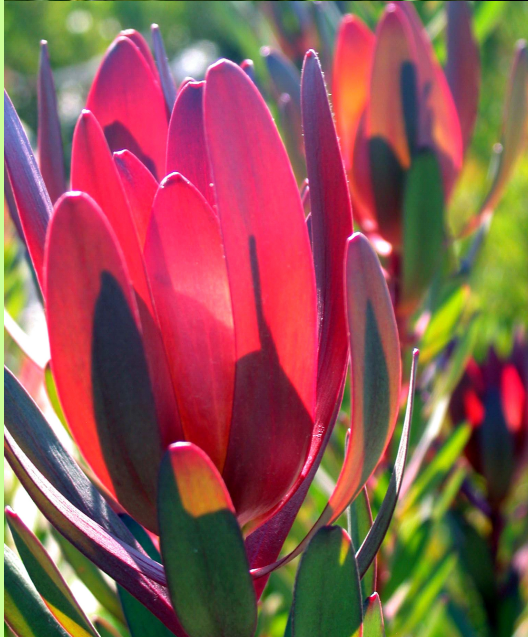


The endosperm works as a barrier actively controlling elemental uptake and preventing access of toxic amounts to embryonic parts. That could explain mechanism of adaptation to hostile environment.



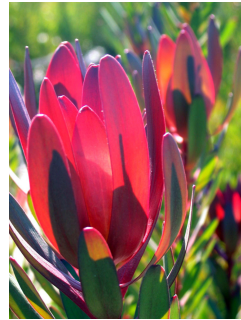
# PROTEA FARMING

## Phosphorus toxicity – a problem for Protea farming on previously agricultural land

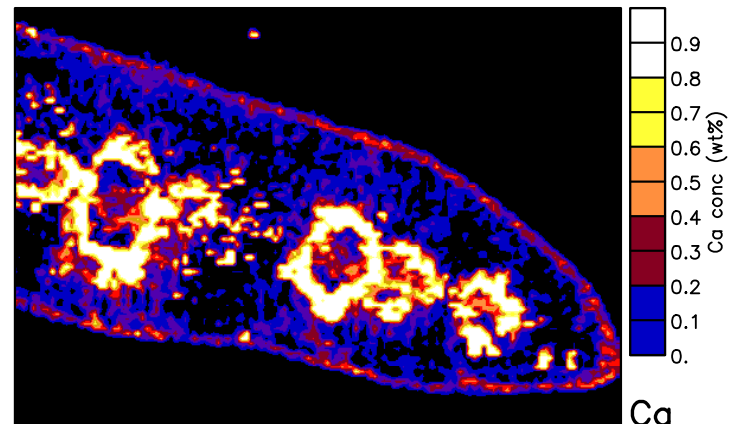
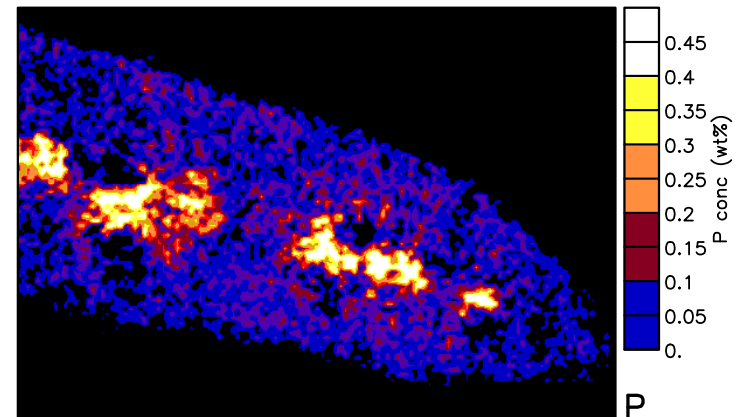
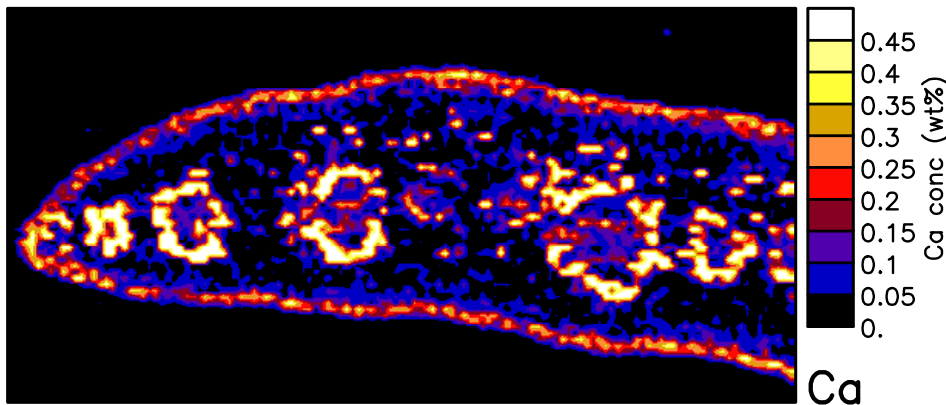
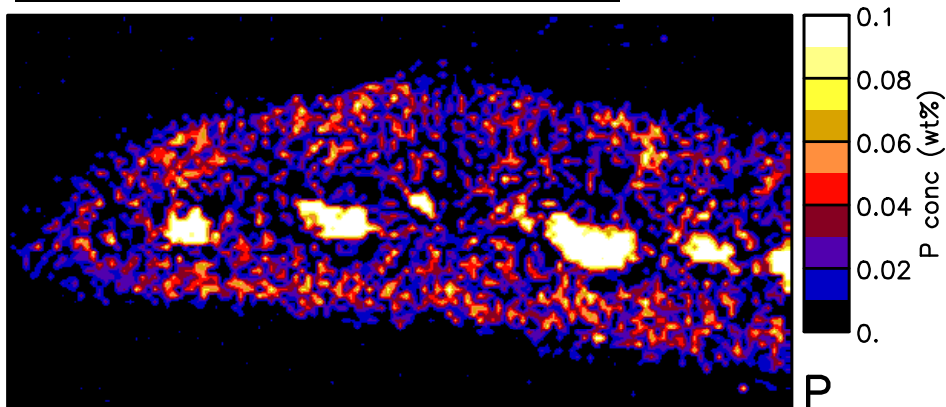
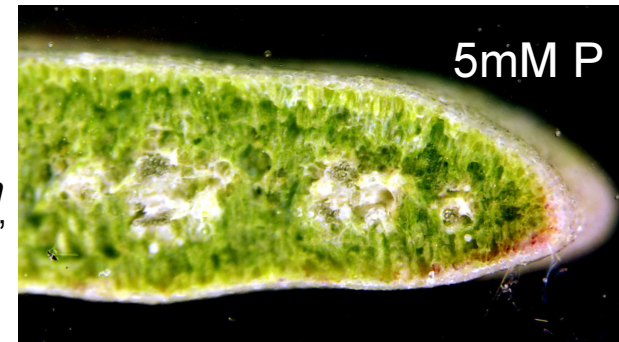


# PROTEA FARMING

Phosphorus toxicity – a problem on previously agricultural land



*Leucadendron*  
'Safari Sunset'



1000

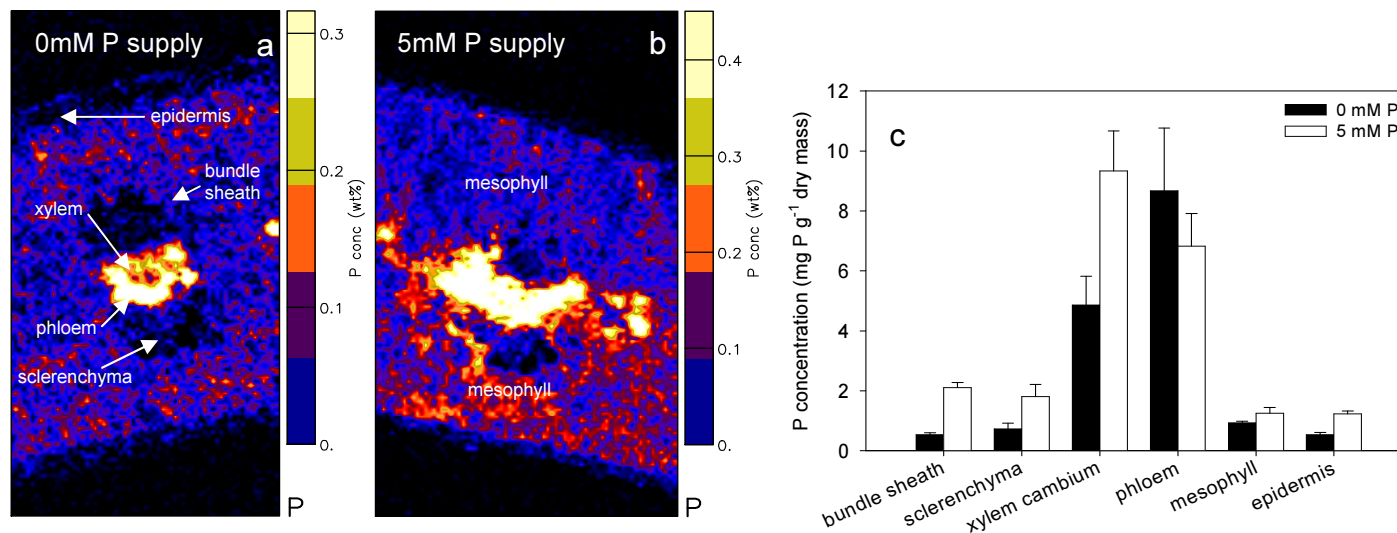
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# Phosphorus toxicity in Proteaceae plants: an association with non-exchangeable binding forms of calcium, iron and manganese in leaf tissues

H.-J. Hawkins<sup>1</sup>, H. Hettasch<sup>2</sup>, J. Mesjasz-Przybylowicz, W.J. Przybylowicz, M. D. Cramer<sup>1</sup>

<sup>1</sup>Department of Botany, University of Cape Town, <sup>2</sup>Arnelia Farms, Hopefield



Location of phosphorus in cross-sections of *Leucadendron* 'Safari Sunset' leaves supplied with no phosphorus (a), high phosphorus (b); and phosphorus concentrations in various leaf tissue types (c).

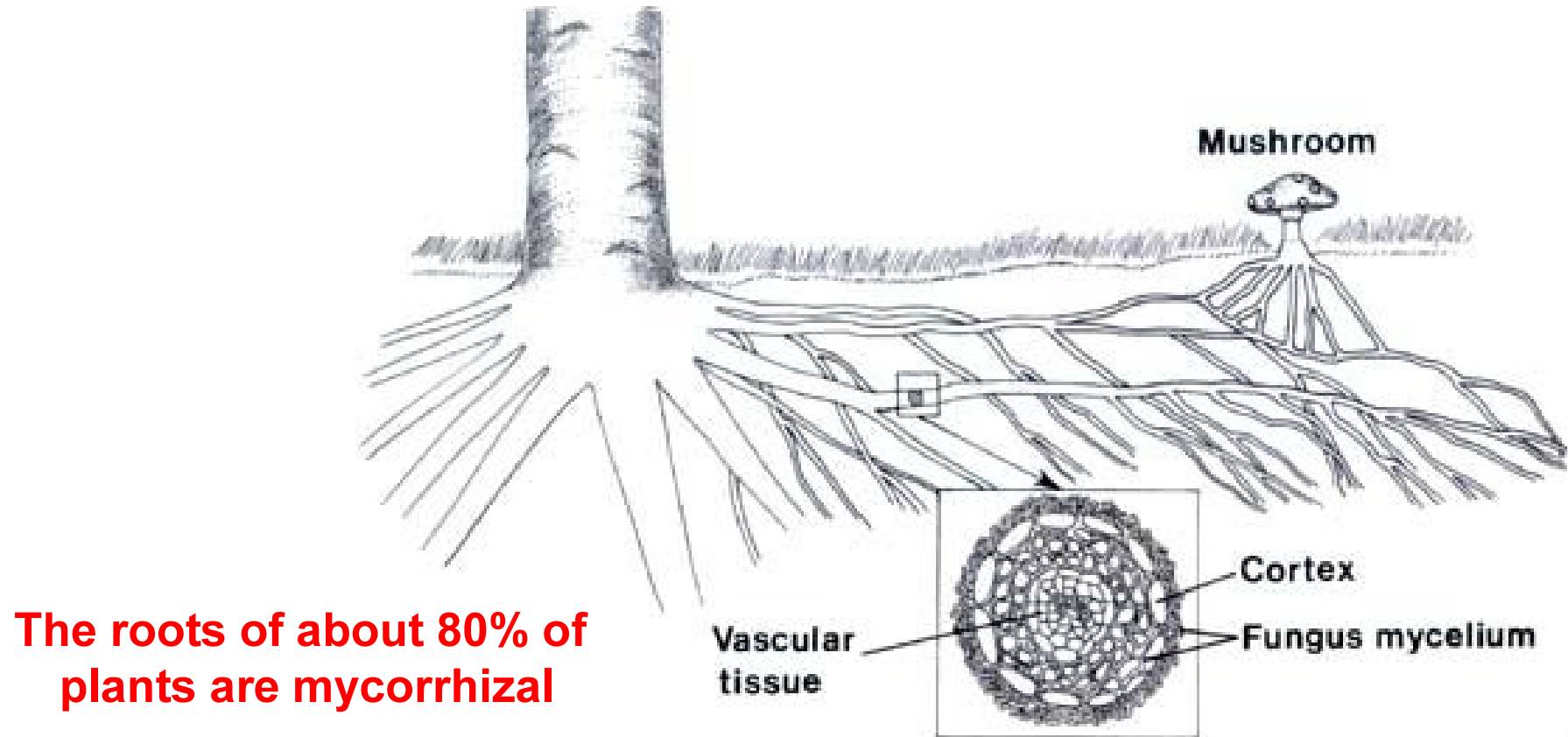
# MYCORRHIZA

From Greek:

**Myco** – “fungus” and **rhiza** – “root”

symbiosis between plant roots and fungi;

modifies the interaction between plants and soil



## **Specific ways in which mycorrhiza assists plants:**

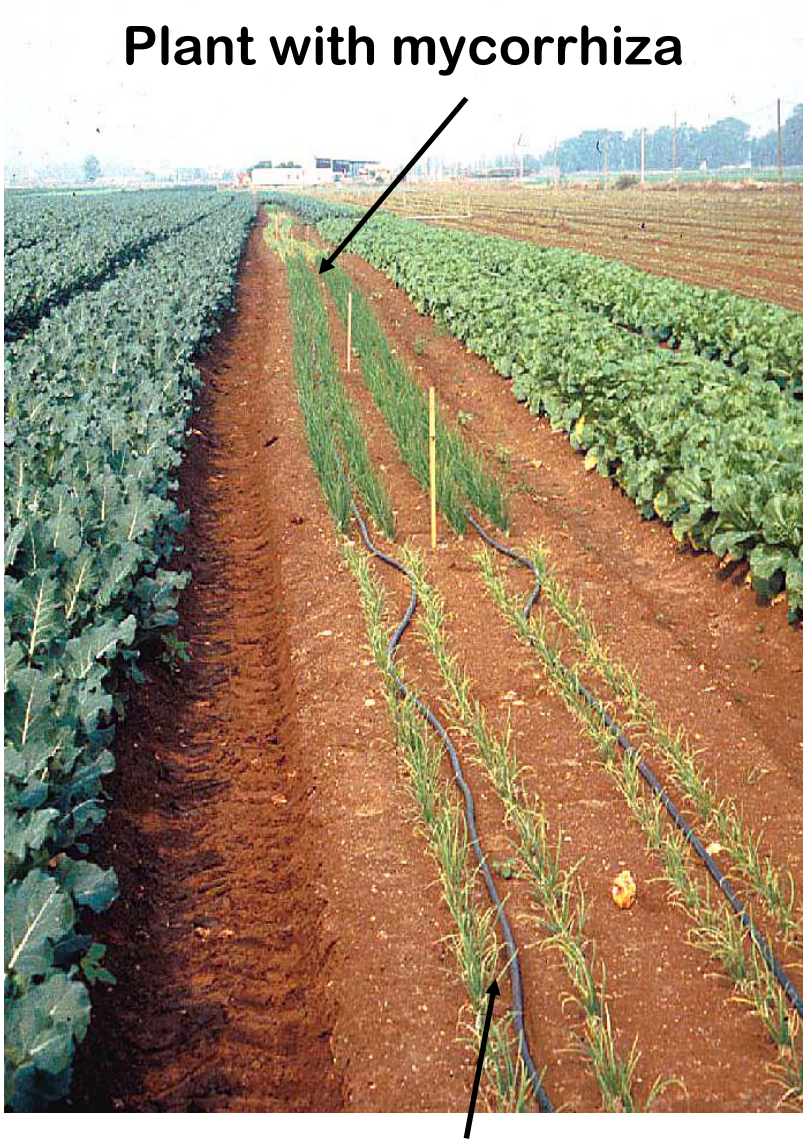
- ☐ **Improves Storage of Carbohydrates**
- ☐ **Improves Mineral Nutrition**
- ☐ **Improves Water Transport**
- ☐ **Can interconnect different host plants**
- ☐ **Assists in heavy metal tolerance**

**Knowledge of elemental concentrations and distribution is essential in studying these processes**

**Mycorrhiza is important in agriculture, forestry, studies of environmental pollution**

## **Mycorrhiza is popular**

**Plant with mycorrhiza**



**Plants without mycorrhiza**



**Maize – always with mycorrhiza**





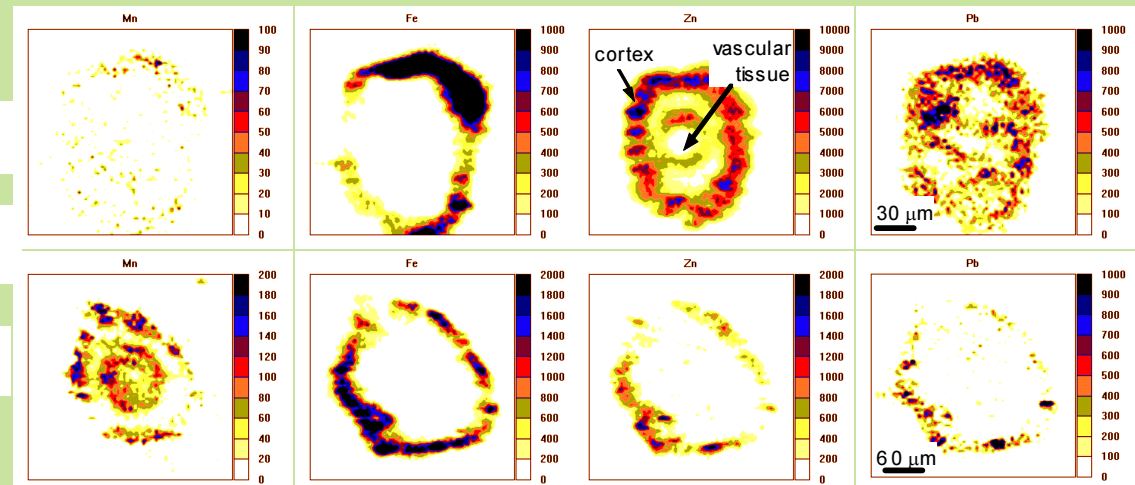
## Mycorrhizal and nonmycorrhizal *Plantago lanceolata* L.

six weeks, greenhouse conditions, rhizo-boxes with industrial waste

mycorrhizal

Quantitative maps of roots

Non-mycorrhizal



different filtering mechanisms of Zn/Pb and Fe/Mn,  
both enable plants to cope with metals present in the environment.

W.J. Przybylowicz, J. Mesjasz-Przybylowicz, P. Migula, M. Nakonieczny, M. Augustyniak, M. Tarnawska, K. Turnau, P. Ryszka, E. Orlowska, S. Zubek, E. Glowacka. X-ray Spectrometry 2005, 34, 285.



# Hyperaccumulation in plants

Certain species accumulate metals to concentrations several orders of magnitude higher than those found in other species on the same site

(above toxicity level for most plants)

**Elements reported to be hyperaccumulated:**

**Al, As, Cd, Co, Cu, Mn, Ni, Pb, Se, Zn**



**Shoots show metal enrichment  
in comparison with roots  
(usually the opposite)**



# Plant-insect herbivore interaction



## *Chrysolina pardalina*

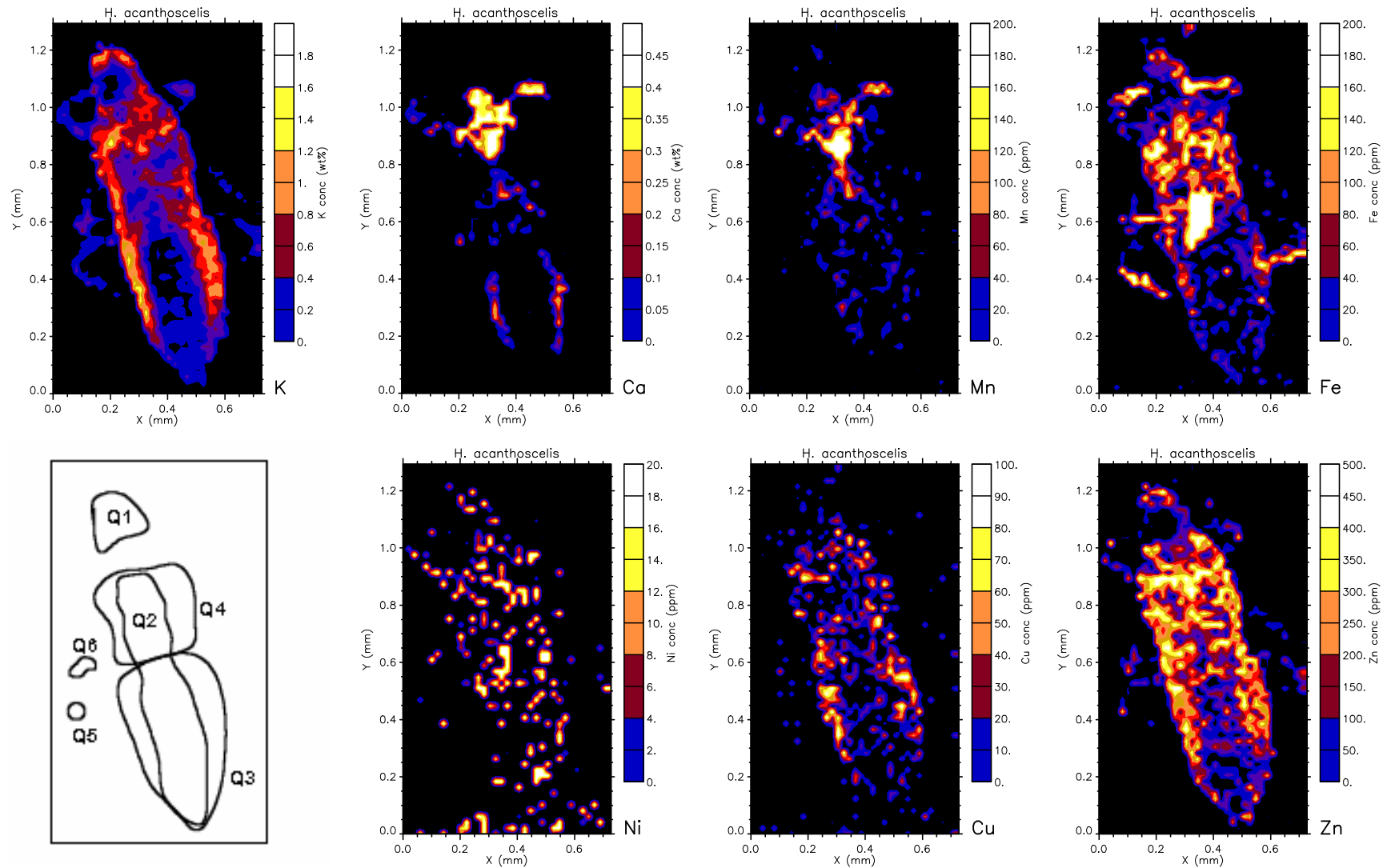
It has been proved that this beetle is able to complete its full life cycle for several generations feeding exclusively on *Berkheya coddii* leaves





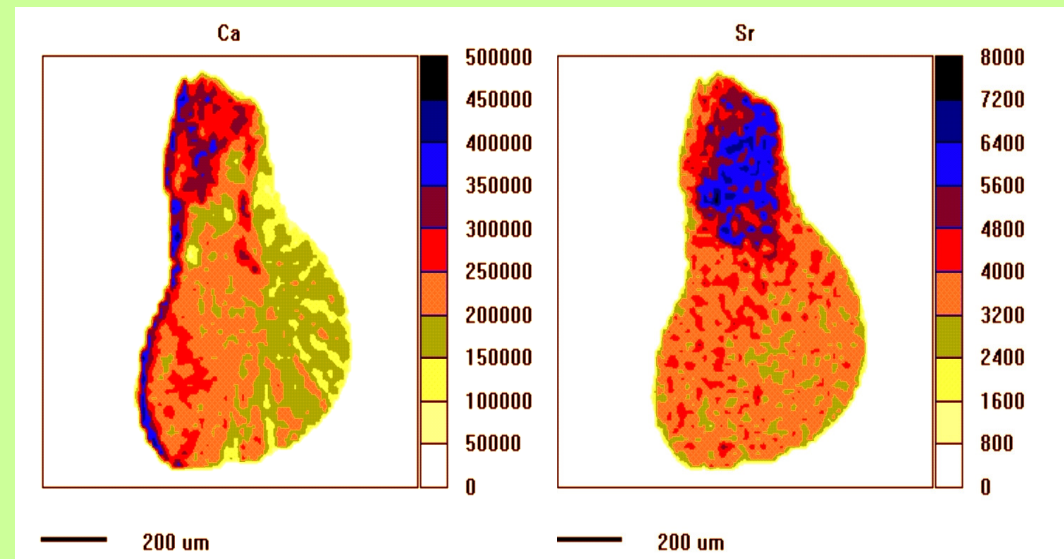
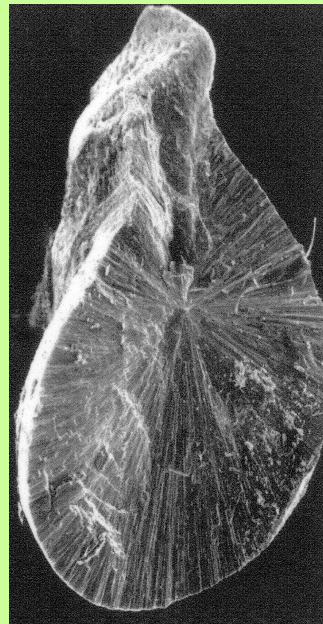
# *Haplothrips acanthoscelis* (Thysanoptera)

Quantitative maps of the whole body and concentrations from marked areas



# Elemental mapping in statoliths of Cephalopods

Statoliths are a part of the balance and movement receptor system



Sr is concentrated in regions adjacent to the macula where the clearest increments are found; this supports the hypothesis linking Sr with the regulation of statolith deposition and the definition of daily increments.

Statolith of squid *Loligo vulgaris reynaudii*

## **Collaborations related to biology, environment and geology**

83 articles in refereed journals  
65 refereed conference proceedings  
258 conference presentations

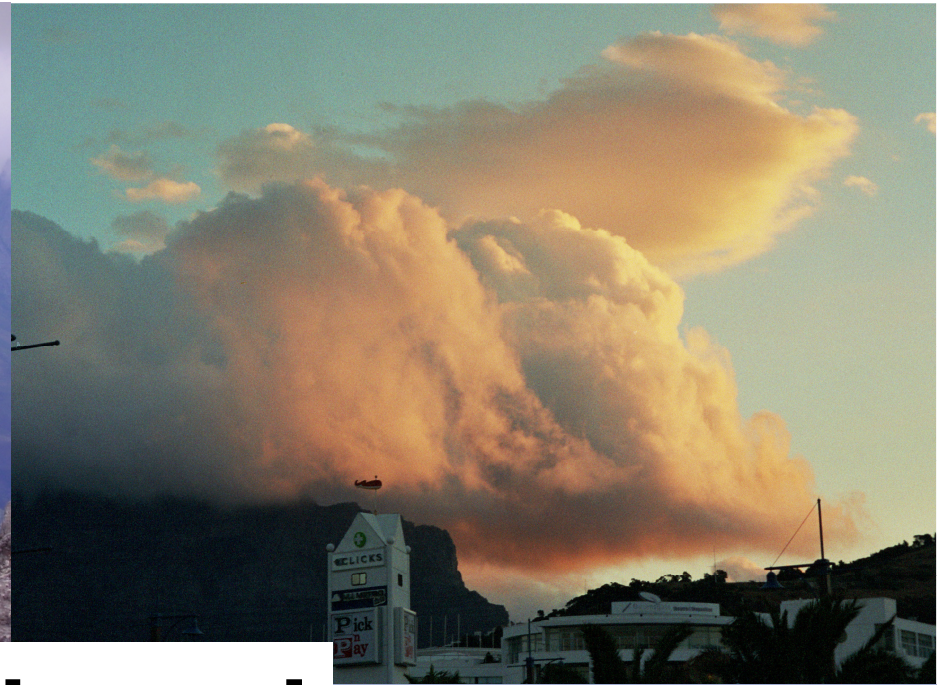
# **Acknowledgments:**

**Accelerator Department: K. Springhorn, S. Marsh, C. Doyle, L.  
Conradie**

**Materials Research Department:  
C. Churms, C. Pineda-Vargas, V. Prozesky, T. Swart, L. Ashworth  
C. Theron**

**All collaborators and contributors**





**Thank you!**

